Synthesis and testosterone 5α -reductase inhibitory activity of 11-substituted 4-aza- 5α -androstane compounds

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Summary — 11α -Acetoxy-, 11α -hydroxy-, 11β -hydroxy-, and 11-oxo-4-aza- 5α -androstane compounds with an N-diphenylmethyl-carbamoyl moiety at the C-17 position were synthesized and their inhibitory activities against rat and human testosterone 5α -reductase were tested. Introduction of the 11α -acetoxy, 11α -hydroxy, 11β -hydroxy and 11-oxo groups into 4-aza- 5α -androstane compounds reduced the inhibitory activity against rat and human 5α -reductase. The 11α -acetoxy-4-aza- 5α -androstane compound in particular lost almost all its activity. However, several compounds with an 11β -hydroxy or 11-oxo group showed inhibitory activities comparable to MK-906. The 4-methyl- 11β -hydroxy-4-aza- 5α -androstane derivative showed the most potent inhibitory activity against rat and human enzyme, and was more active than MK-906.

testosterone 5α -reductase inhibitor/ synthesis/ 4-aza- 5α -androstane/ steroid/ prostatic hypertrophy

Introduction

Benign prostatic hypertrophy is known to be caused by excessive accumulation of dihydrotestosterone, which is formed from testosterone by testosterone 5α reductase. In a previous paper we reported the synthesis and biological activities of new testosterone 5αreductase inhibitors, 4-aza-5α-androstane compounds modified in the B-ring or C-17 carbamoyl group [1]. In that study, we found that 4-aza- 5α -androstane compounds with two aromatic moieties in the C-17 carbamoyl group showed potent inhibitory activities against rat and human 5α-reductase. In the course of our studies on steroidal 5α-reductase inhibitors we became interested in introducing a hydroxy function onto the C-11 position of 4-aza-5α-androstane 1 (fig 1) with an N-diphenylmethylcarbamoyl group at the C-17 position. The importance of the 11B-hydoxy group for steroidal activity is well established [2]. In this paper, we describe a new chemical synthesis of 11-oxygenated 4-aza-5α-androstane derivatives including 11β-hydroxy-4-aza-5α-androstane compound 12 and their 5α -reductase inhibitory activities [3]. The structure—activity relationships are also discussed.

Chemistry

First, a key intermediate, 11α -acetoxy-4-aza-5 α -pregnane-3,20-dione **6**, was synthesized from 11α -hydroxyprogesterone according to a method similar to that described by Rasmusson et al (scheme 1) [4, 5]. Oxidative cleavage of the A ring of 11α -hydroxyprogesterone gave a carboxylic acid **2** (86% yield). Heating of **2** with ammonia, followed by acetylation with acetic anhydride afforded 3-oxo-4-aza-5-ene derivative **4** (53% yield). Hydrogenation of **4** in the presence of a platinum catalyst and then Jones oxidation gave key intermediate **6** (58% yield).

Next, the crucial transformation of the C-17 acetyl group of 6 to a methoxy carbonyl moiety was per-

Fig 1. Compounds 1 and MK-906.

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Scheme 1. (a) KMnO₄, NaIO₄, Na₂CO₃, $tBuOH-H_2O$, reflux; (b) liquid NH₃, ethylene glycol, $0\rightarrow170\,^{\circ}C$; (c) Ac₂O, pyridine, rt; (d) H₂/Pt, AcOH, 60 °C; (e) Jones reagent, acetone, rt; (f) 1.1 eq I₂, pyridine, $50\rightarrow110\,^{\circ}C$ then MeONa, MeOH, reflux.

formed using a modified method to that described by Rasmusson et al (scheme 1) [4]. Treatment of 6 with 1.1 equiv of I_2 in pyridine while the reaction temperature was raised gradually from 50 to 110 °C over 10.5 h, was followed by treatment with NaOMe to give 7 in 35% yield.

Finally, ester 7 was led to the desired products by the following reactions. Hydrolysis of 7 afforded carboxylic acid 8 (61% yield), which was converted to 11-oxygenated derivatives 9–14 (scheme 2). Condensation reaction of 8 with diphenylmethylamine and diethyl phosphorocyanidate [1] afforded the 11 α -hydroxy compound 9 (92% yield). Compound 9 was oxidized to afford the 11-oxo compound 11 (84% yield). Reduction of 11 with NaBH₄ selectively gave the 11 β -hydroxy compound 12 (75% yield). This selectivity due to steric hindrance on the upper side by the 18- and 19-methyl groups is well known in steroid chemistry. Acetylation of the 11 α -hydroxy compound 9 gave the 11 α -acetoxy compound 10 (84% yield). N-

Methylation of 11 gave the 4-methyl-11-oxo compound 13 (61% yield). Compound 13 was reduced to afford the 4-methyl-11 β -hydroxy compound 14 (96% yield).

The synthesis of Δ^{1} -11-oxo derivatives **18** and **19** is shown in scheme 3. Dehydrogenation of **7** with bistrimethylsilyltrifluoroacetamide and DDQ gave the Δ^{1} -ester **15** (67% yield) [6]. Oxidation of **15** with a Jones reagent followed by hydrolysis afforded the Δ^{1} -11-oxo carboxylic acid **17** (71% yield). Amidation of **17** with diphenylmethylamine and diethyl phosphorocyanidate gave the Δ^{1} -11-oxo derivative **18** (68% yield). *N*-Methylation of **18** with potassium *t*-butoxide and iodomethane gave the Δ^{1} -4-methyl-11- oxo compound **19** (87% yield).

Results and discussion

The testosterone 5α -reductase inhibitory activities of the synthesized compounds are listed in table I. The 11α -acetoxy derivative **10**, 11-oxo derivative **11**, 11α hydroxy derivative 9 and 11β -hydroxy derivative 12 showed relatively low inhibitory rates in rat 5α-reductase tests compared with the original 11-unsubstituted compound 1 [1]. The activities in descending order are 12, 9, 11 and 10. The 11α -acetoxy compound 10 had almost no activity. The 11β-hydroxy compound 12 showed potent inhibitory activity comparable to MK906 (finasteride, a drug for benign prostatic hypertrophy) [1, 5] against rat and human 5α -reductase. Further modification of 11 and 12 afforded 4-methyl compounds 13 and 14, which showed increased inhibitory activities more active than MK-906 against rat and human enzyme. Introduction of a double bond at the C-1 position in 11 and 13 gave derivatives 18 and 19. Compounds 18 and 19 showed more potent inhibitory activities than MK-906 against rat enzyme. However, compound 18 was very weakly active against human 5α-reductase. In contrast, compound 19 retained its inhibitory activity against human enzyme.

In conclusion, 11α -acetoxy-, 11α -hydroxy-, 11β -hydroxy-, and 11-oxo-4-aza- 5α -androstane compounds with an *N*-diphenylmethylcarbamoyl moiety at the C-17 position were synthesized and their inhibitory activities against rat and human testosterone 5α -reductase tested. Introduction of 11α -acetoxy, 11α -hydroxy, 11β -hydroxy, and 11-oxo groups into compound 1 reduced the inhibitory activity against rat and human 5α -reductase. The 11α -acetoxy compound 10 lost almost all its activity. However, compounds 12–14, 18, and 19 showed potent inhibitory activities comparable to MK-906. 4-Methyl- 11β -hydroxy-4-aza- 5α -androstane derivative 14 showed the most

Scheme 2. (a) KOH, MeOH/H₂O, reflux; (b) Ph₂CHNH₂, DEPC, Et₃N, CH₂Cl₂, rt; (c) Ac₂O, pyridine, rt; (d) Jones reagent, acetone, rt; (e) NaBH₄, EtOH/H₂O, rt; (g) *t*-BuOK, MeI, *t*-BuOH, rt.

potent inhibitory activity against rat and human enzyme; it was more active than MK-906 and as active as compound 1.

Experimental protocols

Chemistry

Melting points were determined with a Yanagimoto micro melting point apparatus MP-500D and are uncorrected. $^1\mathrm{H-NMR}$ spectra were recorded on a Jeol JNM-GX270 or Jeol JNM-EX270 spectrometer (270 MHz). Chemical shifts are given in ppm (δ) using tetramethylsilane as an internal standard. IR spectra were measured with a Nic.5SXC, Jasco A-302, Jasco FTIR8300, Jasco FTIR8900, or Jasco A-102 spectrometer. Mass spectra were measured with a Jeol JMS-D300, JMS-AX505H, or JMS-AX505W spectrometer. Thin-layer chromatography (TLC) was run on silica gel-coated plates (E Merck, Silica gel 60 F_{254} precoated) of thickness 0.25 mm. Preparative

thin-layer chromatography was carried out on similar plates with a thickness of 2 mm. Silica gel 60 (E Merck, 70–230 mesh) was used for column chromatography.

5,20-Dioxo-11α-hydroxy-A-nor-3,5-secopregnan-3-oic acid 2 A solution of 11α-hydroxy-3,20-dioxopregn-4-ene (10.5 g, 31.8 mmol) and Na₂CO₃ (4.40 g, 41.5 mmol) in a mixture of 2-methyl-2-propanol (150 mL) and water (20 mL) was brought to reflux. A solution of KMnO₄ (0.34 g, 2.15 mmol) and NaIO₄ (41.0 g, 192 mmol) in hot water (200 mL) was added dropwise over 30 min and the mixture refluxed for 1 h. The reaction mixture was cooled to room temperature and the formed solid was filtered and washed with water. After evaporation of 2-methyl-2-propanol from the filtrate, the remaining water solution was acidified with 1 N HCl. The precipitate was filtered, washed with water, and dried under air to give 2 (9.54 g, 86%), mp 101–104 °C. ¹H-NMR (pyridine- d_5) δ : 0.73 (3H, s), 1.12–1.30 (3H, m), 1.36 (3H, s), 1.52–1.82 (6H, m), 2.09 (3H, s), 2.37 (1H, m), 2.45-2.63 (4H, m), 2.77 (1H, m), 2.92-3.05 (2H, m), 3.12 (1H, m), 4.27 (1H, dt, J = 5, 10 Hz). IR (KBr): 3483, 2976, 2951, 2889, 1726, 1688 cm⁻¹. HR-MS m/z: calc for C₂₀H₃₀O₅ (M+): 350.2094; found: 350.2100.

Scheme 3. (a) DDQ, N,O-bistrimethylsilyltrifluoracetamide, dioxane, rt to reflux; (b) Jones reagent, acetone, rt; (c) KOH, dioxane/H₂O, reflux; (d) Ph₂CHNH₂, DEPC; Et₃N, CH₂Cl₂, rt; (e) t-BuOK, MeI, t-BuOH, rt.

3,20-Dioxo-11α-hydroxy-4-azapregn-5-ene 3

Liquid NH₃ (20.0 mL) was added to a suspension of **2** (9.50 g, 27.1 mmol) in ethylene glycol (65 mL) at 0 °C. The whole was gradually heated to 170 °C over 1 h and stirred for 45 min at the same temperature. The reaction mixture was cooled to room temperature, acidified with 1 N HCl and extracted with CH₂Cl₂. The combined organic layer was washed with brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with CH₂Cl₂/acetone (3:1 to 2:3) was evaporated and the residue was triturated with Et₂O to give **3** (7.16 g, 80%), mp 189–193 °C. ¹H-NMR (pyridine- d_5) δ : 0.72 (3H, s), 1.08–1.84 (9H, m), 1.35 (3H, m), 1.98–2.15 (1H, m), 2.09 (3H, s), 2.36 (1H, m), 2.52–2.82 (4H, m), 3.41 (1H, m), 4.28 (1H, m), 5.23 (1H, dd, J = 2, 5 Hz), 5.84 (1H, d, J = 6 Hz), 10.31 (1H, s). IR (KBr): 3400, 2941, 2920, 1703, 1690, 1659 cm⁻¹. HR-MS m/z: calc for C₂₀H₂₉NO₃ (M⁺): 331.2148; found: 331.2131.

11α-Acetoxy-3,20-dioxo-4-azapregn-5-ene 4

A solution of 3 (2.28 g, 6.88 mmol) in a mixture of Ac_2O (10.0 mL) and pyridine (20.0 mL) was stirred at room temperature for 3 h. After addition of ice, the reaction mixture was extracted with CH_2Cl_2 . The combined organic layer was washed with 1 N HCl and brine. The organic layer was dried over Na_2SO_4 and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with CH_2Cl_2 /acetone (4:1 to 2:3) was evaporated and the residue was triturated with Et_2O to give 4 (1.70 g, 66%), mp 199–204 °C. ¹H-NMR (CDCl₃) δ : 0.74 (3H, s), 1.15–2.63 (16H, m), 1.18 (3H, s), 2.06 (3H, s), 2.12 (3H, s), 4.90 (1H, dd, J=2, 5 Hz), 5.29 (1H, dt, J=5, 10 Hz), 7.32 (1H, br). IR (KBr): 3182, 3065, 2980, 2922, 2886, 1735, 1703, 1664 cm⁻¹. HR-MS m/z: calc for $C_{22}H_{31}NO_4$ (M+): 373.2253; found: 373.2244.

11α-Acetoxy-20-hydroxy-3-oxo-4-aza-5α-pregnane 5

A suspension of 4 (904 mg, 2.42 mmol) and platinum dioxide (300 mg) in acetic acid (20 mL) was stirred under hydrogen atmosphere at 60 °C for 2 h. The platinum catalyst was filtered off and washed with MeOH. The filtrate was evaporated and the residue was chromatographed on a silica-gel column. The fraction eluted with CH₂Cl₂/acetone (3:2 to 3:7) was evaporated to give 5 (697 mg, 76%) as a foam. $^1\text{H-NMR}$ (CDCl₃) δ : 0.70 and 0.82 (total 3H, each s), 0.99 and 1.00 (total 3H, each s), 1.07–1.88 (18H, m), 1.99 and 2.03 (total 3H, each s), 2.31–2.58 (3H, m), 3.13 and 3.17 (total 1H, each m), 3.71 (1H, m), 5.19 (1H, m), 5.95 and 6.09 (total 1H, each br). IR (KBr): 3210, 2967, 2943, 2871, 1731, 1707, 1666 cm⁻¹. HR-MS m/z: calc for $C_{22}H_{36}NO_4$ ((M+1)+): 378.2644; found: 378.2667.

11α -Acetoxy-3,20-dioxo-4-aza-5 α -pregnane 6

Jones reagent (4.00 mL) was added to a solution of 5 (4.60 g, 12.2 mmol) in acetone (50 mL) at room temperature and the mixture was stirred for 30 min. Additional Jones reagent (2.00 mL) was added and the mixture stirred for a further 1 h. After addition of 2-propanol, the reaction mixture was diluted with brine and extracted with CH2Cl2. The combined organic layer was washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and evaporated to dryness. The residue was chromatographed on a silica-gel column. The fraction eluted with CH₂Cl₂/acetone (3:1 to 1:1) was evaporated and the residue was triturated with Et₂O to give **6** (3.50 g, 76%), mp 178–183 °C. ¹H-NMR (CDCl₃) δ : 0.70 (3H, s), 0.82–2.65 (18H, m), 0.99 (3H, s), 2.04 (3H, s), 2.11 (3H, s), 3.14 (1H, dd, J = 4, 12 Hz), 5.19 (1H, dt, J = 4, 12 Hz)J = 5, 11 Hz), 6.19 (1H, br). IR (CHCl₃): 2960, 2870, 1725, 1700, 1655 cm⁻¹. HR-MS m/z: calc for $C_{22}H_{33}NO_4$ (M+): 375.2409; found: 375.2385.

Table I. Inhibitory activities of 11-substituted 4-aza-5α-androstane compounds.

Compound	R_I	R_2	R_3	R_4	Rat 50-reductase % inhibition at 10 ⁻⁸ M	Human 5α -reductase relative inhibitory potency ^a to MK-906 (MK-906 = 1)
9	Н	ОН	Н	H	37	<0.1
11	O		Н	Н	17	0.30
12	ОН	Н	Н	Н	50	0.55
10	Н	OAc	Н	Н	6.8	<0.1
13	O		Me	Н	74	1.6
14	OH	Н	Me	Н	74	2.9
18	O		H	_b	39	< 0.1
19	O		Me	_b	33	1.0
1	Н	Н	Н	H	89	5.6
MK-906					28	1.0

^aThe relative inhibitory potency was calculated from the inhibition curve of MK-906 (0.25 × 10-8 M to 8 × 10-8 M) and the % inhibition of compounds at 10-7 M or 10-8 M. The % inhibition of MK-906 at 10-8 M varied from 31 to 65% for human 5α -reductase from BPH patients. b— means a single bond combined with both R_4 .

Methyl 11 α -hydroxy-3-oxo-4-aza-5 α -androstane-17 β -carboxy-late 7

A solution of a mixture of 6 (10.2 g, 27.2 mmol) and I_2 (7.90 g, 31.1 mmol) in pyridine (200 mL) was heated from 50 to 110 °C over 10.5 h and stirred at the same temperature for 6 h under N₂ atmosphere. The reaction mixture was cooled to room temperature and absolute MeOH (250 mL) and 28% MeONa in MeOH solution (110 ml, 540 mmol) were added. The mixture was stirred at 65 °C for 1 h. 28% MeONa in MeOH solution (100 mL, 490 mmol) was added and the mixture stirred for an additional 1 h. The reaction mixture was neutralized with 1 N HCl at 0 °C and the MeOH evaporated. The residue was diluted with 1 N HCl and extracted with CHCl3. The combined organic layer was washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silicagel column. The fraction eluted with 2-propanol/toluene/acetone (3:37:60 to 5:35:60) was evaporated and the residue was triturated with Et_2O to give 7 (3.34 g, 35%), mp 248–251 °C. ¹H-NMR (CDCl₃) δ : 0.69 (3H, s), 0.88–2.48 (17H, m), 1.02 (3H, s), 2.68 (1H, m), 3.14 (1H, dd, J = 4, 12 Hz), 3.69 (3H, s), 3.96 (1H, dt, J = 5, 10 Hz), 6.19 (1H, br). IR (KBr): 3396, 2940, 2876, 1733, 1657 cm⁻¹. HR-MS m/z: calc for C₂₀H₃₁NO₄ (M+): 349.2253; found: 349.2239.

11 α-Hydroxy-3-oxo-4-aza-5α-androstane-17 β -carboxylic acid 8 A solution of 7 (102 mg, 0.29 mmol) in a mixture of MeOH (2.0 mL) and 10% KOH aqueous solution (1.0 mL) was refluxed under N_2 atmosphere for 4 h. After evaporation of

MeOH, the residue was acidified with 1 N HCl. The precipitate was filtered and washed with water to give **8** (60 mg, 61%), mp 245 °C (dec). ¹H-NMR (pyridine- d_s) δ: 0.85–2.02 (12H, m), 0.97 (3H, s), 1.11 (3H, s), 2.40–2.70 (4H, m), 2.97 (1H, dd, J = 4, 12 Hz), 3.13–3.29 (2H, m), 4.21 (1H, br), 5.56 (1H, br). IR (KBr): 3420, 3269, 3177, 2966, 2935, 1699, 1632 cm⁻¹. HR-MS m/z: calc for C₁₉H₂₉NO₄ (M+): 335.2096; found: 335.2107.

N-Diphenylmethyl- 11α -hydroxy-3-oxo-4-aza- 5α -androstan- 17β -carboxamide **9**

A solution of 8 (40 mg, 0.12 mmol), diphenylmethylamine (80 μ L, 0.46 mmol), diethyl phosphorocyanidate (70 μ L, 0.46 mmol), and triethylamine (55 μ L, 0.40 mmol) in CH₂Cl₂ (2.0 mL) was stirred at room temperature for 24 h. The reaction mixture was diluted with 1 N HCl and extracted with CH₂Cl₂. The combined organic layer was washed with saturated aqueous NaHCO3 solution and brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with 2-propanol/CH₂Cl₂/acetone (3:37:60 to 1:3:6) was evaporated and the residue was crystallized from EtOAc to give 9 (55 mg, 92%), mp 160–163 °C. ¹H-NMR (CDCl₃) δ : 0.70 (3H, s), 0.84–1.95 (14H, m), 1.01 (3H, s), 2.15–2.30 (2H, m), 2.44 (1H, m), 2.68 (1H, m), 3.15 (1H, dd, J = 5, 12 Hz), 3.98 (1H, dt, J =4, 10 Hz), 5.88 (1H, d, J = 8 Hz), 6.28 (1H, d, J = 8 Hz), 6.48 (1H, br), 7.19-7.38 (10H, m). IR (KBr): 3304, 2931, 2873, 1656 cm⁻¹. HR-MS m/z: calc for $C_{32}H_{40}N_2O_3$ (M+): 500.3039; found: 500.3068. Anal calc for C₃₂H₄₀N₂O₃·3/4H₂O: C, 74.75; H. 8.14; N. 5.45; found: C. 74.47; H. 7.91; N. 5.31.

N-Diphenylmethyl-11 α -acetoxy-3-oxo-4-aza-5 α -androstane-17 β -carboxamide **10**

A solution of **9** (52 mg, 0.10 mmol) in a mixture of pyridine (1.0 mL) and Ac₂O (0.5 mL) was stirred at room temperature for 12 h. After ice was added to the reaction mixture, the solution was diluted with 1 N HCl and extracted with CH₂Cl₂. The combined organic layer was washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with CH₂Cl₂/acetone (7:3 to 4:6) was evaporated and the residue was crystallized from a mixture of CH₂Cl₂/EtOAc to give **10** (47 mg, 84%), mp 258–261 °C. ¹H-NMR (CDCl₃) δ : 0.72 (3H, s), 0.96–1.90 (14H, m), 0.98 (3H, s), 2.00 (3H, s), 2.15–2.43 (4H, m), 3.13 (1H, dd, J = 4, 12 Hz), 5.16 (1H, dt, J = 5, 10 Hz), 5.64 (1H, br), 5.84 (1H, d, J = 8 Hz), 6.27 (1H, d, J = 8 Hz), 7.19–7.39 (10H, m). IR (KBr): 3306, 2969, 2936, 1731, 1664 cm⁻¹. HR-MS m/z: calc for C₃₄H₄₂N₂O₄·2/3H₂O: C, 73.62; H, 7.87; N, 5.05. Found: C, 73.57; H, 8.00; N, 4.97.

N-Diphenylmethyl-3,11-dioxo-4-aza-5 α -androstane-17 β -carboxamide 11

Jones reagent (0.20 mL) was added to a solution of 9 (208 mg, 0.42 mmol) in acetone (15 mL) at room temperature and the mixture was stirred for 1 h. After addition of 2-propanol, the reaction mixture was made basic with 1 N NaOH and diluted with CH2Cl2. The solution was passed through celite and the filtrate extracted with CH₂Cl₂. The combined organic layer was washed with brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silicagel column. The fraction eluted with 2-propanol/CH₂Cl₂/acetone (0:1:1 to 2:38:60) was evaporated and the residue crystallized from a mixture of CH₂Cl₂/EtOAc to give 11 (171 mg, 83%), mp 279 °C (dec). ¹H-NMR (CDCI₃) δ: 0.65 (3H, s), 1.09 (3H, s), 1.17–2.03 (10H, m), 2.17–2.65 (8H, m), 3.02 (1H, dd, J = 4, 11 Hz), 5.72 (1H, br), 5.85 (1H, d, J = 8 Hz), 6.25 (1H, d, J = 8 Hz), 7.19-7.39 (10H, m). IR (KBr): 3305, 2943, 2874, 1706, 1657 cm⁻¹. HR-MS m/z: calc for $C_{32}H_{38}N_2O_3$ (M+): 498.2883; found: 498.2876. Anal calc for $C_{32}H_{38}N_2O_3\cdot 3/4H_2O$: C, 75.04; H, 7.77; N, 5.47. Found: C, 74.93; H, 7.86; N, 5.43.

N-Diphenylmethyl-11β-hydroxy-3-oxo-4-aza-5α-androstane-17β-carboxamide 12

NaBH₄ (77 mg, 2.00 mmol) was added to a solution of 11 (102 mg, 0.20 mmol) in a mixture of EtOH (6.0 mL) and water (0.50 mL) and the mixture was stirred at room temperature for 24 h. The reaction mixture was cooled to 0 °C and AcOH was added. The solution was diluted with water and extracted with CH₂Cl₂. The combined organic layer was washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with CH₂Cl₂/acetone (1:1 to 4:6) was evaporated and the residue was triturated with Et₂O to give 12 (76 mg, 75%), mp 153–156 °C. ¹H-NMR (CDCl₃) δ : 0.77–2.22 (16H, m), 0.91 (3H, s), 1.12 (3H, s), 2.40–2.49 (2H, m), 3.03 (1H, dd, J = 4, 11 Hz), 4.30 (1H, d, J = 2 Hz), 5.58 (1H, br), 5.89 (1H, d, J = 8 Hz), 6.28 (1H, d, J = 8 Hz), 7.21-7.38 (10H, m). IR (KBr): 3309, 2931, 2871, 1652 cm⁻¹. HR-MS m/z: calc for $C_{32}H_{40}N_2O_3$ (M⁺): 500.3039; found: 500.3063. Anal calc for $C_{32}H_{40}N_2O_3 \cdot 3/4H_2O$: C, 74.75; H, 8.14; N, 5.45. Found: C, 74.79; H, 8.34; N, 5.21.

N-Diphenylmethyl-3,11-dioxo-4-methyl-4-aza-5 α -androstan-17 β -carboxamide 13

t-BuOK (34 mg, 0.30 mmol) was added to a solution of 11 (60 mg, 0.12 mol) in 2-methyl-2-propanol (4.0 mL) and the

mixture was stirred at room temperature for 1 h under N₂ atmosphere. Methyl iodide (0.12 mL, 1.93 mmol) was then added to the reaction mixture and stirring continued for 6 h. The reaction mixture was diluted with 1 N HCl and extracted with CH2Cl2. The combined organic layer was washed with saturated aqueous NaHCO₃ solution and brine. The organic layer was dried over Na₂SO₄ and evaporated. The residue was purified by preparative thin-layer chromatography developed with CH₂Cl₂/acetone (1:1). A part showing an R_f-value of 0.5 was extracted with acetone. After evaporation of the acetone, the residue was crystallized from a mixture of CH₂Cl₂/ EtOAc to give 13 (38 mg, 61%), mp 142-145 °C. 1H-NMR $(CDCl_3)$ δ : 0.65 (3H, s), 1.07–2.65 (18H, m), 1.08 (3H, s), 2.92 (3H, s), 3.00 (1H, dd, J = 3, 12 Hz), 5.84 (1H, d, J = 8 Hz), 6.25 (1H, d, J = 8 Hz), 7.20–7.39 (10H, m). IR (KBr): 3308, 2959, 2876, 1706, 1645 cm⁻¹. HR-MS m/z: calc for $C_{33}H_{40}N_2O_3$ (M+): 542.3039; found: 512.3011. Anal calc for $C_{33}H_{40}N_2O_{3}$ 3/4H₂O: C, 75.32; H, 7.95; N, 5.32. Found: C, 75.40; H, 8.21; N, 5.30.

N-Diphenylmethyl-11 β -hydroxy-4-methyl-3-oxo-4-aza-5 α -androstane-17 β -carboxamide **14**

Compound **14** (cryst, CH₂Cl₂/EtOAc) was synthesized in 96% yield from **13** using a method similar to that described for the preparation of **12**. Mp 164–167 °C. ¹H-NMR (CDCl₃) δ : 0.78–2.23 (16H, m), 0.91 (3H, s), 1.10 (3H, s), 2.50–2.58 (2H, m), 2.94 (3H, s), 2.98 (1H, dd, J = 3, 12 Hz), 4.31 (1H, d, J = 3 Hz), 5.87 (1H, d, J = 8 Hz), 6.29 (1H, d, J = 8 Hz), 7.20–7.38 (10H, m). IR (KBr): 3441, 3320, 2943, 2874, 1623 cm⁻¹. HR-MS m/z: calc for C₃₃H₄₂N₂O₃ (M+): 514.3196; found: 514.3176. Anal calc for C₃₃H₄₂N₂O₃·1/2H₂O: C, 75.68; H, 8.28; N, 5.35. Found: C, 75.83; H, 8.04; N, 5.26.

Methyl $II\alpha$ -hydroxy-3-oxo-4-aza- 5α -androst-1-ene- $I7\beta$ -carboxylate **15**

Dichlorodicyanohydroquinone (240 mg, 1.06 mmol) and N,Obistrimethylsilyltrifluoroacetamide (1.45 mL, 5.50 mmol) were added to a solution of 7 (316 mg, 0.90 mmol) in dry dioxane (10 mL). The mixture was stirred at room temperature for 5 h under N₂ atmosphere and refluxed for 18 h. The reaction mixture was cooled to room temperature, and then CH2Cl2 (8.0 mL) and 1% aqueous NaHSO₃ solution (2.0 mL) were added. The precipitated solid was filtered off and washed with CH₂Cl₂. The filtrate was diluted with 10% aqueous HCl solution and extracted with CHCl₃. The combined organic layer was washed with 1 N HCl and then brine. The organic layer was dried over Na2SO4 and evaporated. The residue was chromatographed on a silica-gel column. The fraction eluted with 2-propanol/CH₂Cl₂/acetone (0:1:1 to 1:39:60) was evaporated and the residue was triturated with Et₂O to give 15 (210 mg, 67%), mp 260 °C (dec). ¹H-NMR (CDCl₃) δ : 0.70 (3H, s), 1.05–1.94 (H, m), 2.16 (1H, m), 2.31 (1H, dd, J = 4, 12 Hz), 2.41 (1H, t, J = 10 Hz), 3.38 (1H, t, J = 8 Hz), 3.69 (3H, s), 4.05 (1H, dt, J = 5, 10 Hz), 5.34 (1H, br), 5.74 (1H, dd, J = 2,10 Hz), 7.95 (1H, d, J = 10 Hz). IR (CHCl₃): 3405, 3340, 2970, 2935, 1718, 1660 cm⁻¹. HR-MS m/z: calc for $C_{20}H_{29}NO_4$ (M⁺): 347.2097; found: 347.2092.

Methyl 3,11-dioxo-4-aza-5α-androst-1-ene-17β-carboxylate 16 Compound 16 was synthesized in 75% yield from 15 using a method similar to that described for the preparation of 11. Mp 284–287 °C. ¹H-NMR (CDCl₃) δ: 0.63 (3H, s), 1.18–2.03 (10H, m), 1.19 (3H, s), 2.18–2.42 (2H, m), 2.57–2.72 (2H, m), 3.30 (1H, dd, J = 7, 9 Hz), 3.69 (3H, s), 5.40 (1H, br), 5.80 (1H, d, J = 10 Hz), 7.18 (1H, d, J = 10 Hz). IR (KBr): 3177, 3102, 2933, 1717, 1703, 1679, 1603 cm⁻¹. HR-MS m/z: calc for $C_{20}H_{27}NO_4$ (M+): 345.1940; found: 345.1944.

3,11-Dioxo-4-aza-5α-androst-1-ene-17β-carboxylic acid 17 A solution of **16** (65 mg, 0.19 mmol) in a mixture of dioxane (3.0 mL) and 10% KOH aqueous solution (1.5 mL) was refluxed for 3 h under N_2 atmosphere. After evaporation of the dioxane, the residue was acidified with 1 N HCl. The precipitate was filtered and washed with water to give **17** (58 mg, 94%), mp >300 °C. ¹H-NMR (CDCl₃ + CD₃OD) δ: 0.69 (3H, s), 1.17–2.77 (14H, m), 1.18 (3H, s), 3.29 (1H, t, J = 8 Hz), 5.77 (1H, d, J = 10 Hz), 7.20 (1H, d, J = 10 Hz). IR (KBr): 3309, 2951, 2867, 2525, 1702, 1648, 1593 cm⁻¹. HR-MS m/z: calc for $C_{10}H_{25}NO_4$ (M+): 331.1783; found: 331.1788.

N-Diphenylmethyl-3,11-dioxo-4-aza-5 α -androst-1-ene-17 β -carboxamide 18

Compound **18** (cryst, CH₂Cl₂/EtOAc) was synthesized in 69% yield from **17** by a similar method to that described for the preparation of **9**. Mp 209–212 °C. ¹H-NMR (CDCl₃) δ : 0.65 (3H, s), 1.17–2.03 (10H, m), 1.18 (3H, s), 2.18–2.50 (3H, m), 2.61 (1H, d, J = 12 Hz), 3.29 (1H, t, J = 8 Hz), 5.36 (1H, br), 5.79 (1H, d, J = 10 Hz), 5.86 (1H, d, J = 8 Hz), 6.25 (1H, d, J = 8 Hz), 7.16 (1H, d, J = 10 Hz), 7.20–7.39 (10H, m). IR (KBr): 3310, 2940, 1706, 1675, 1600 cm⁻¹. HR-MS m/z: calc for $C_{32}H_{36}N_2O_3$ (M+): 496.2726; found: 496.2743. Anal calc for $C_{32}H_{36}N_2O_3$ •5/4H₂O: C, 74.03; H, 7.47; N, 5.40. Found: C, 73.86; H, 7.23; N, 5.47.

N-Diphenylmethyl-3,11-dioxo-4-methyl-4-aza-5α-androst-1-ene-17β-carboxamide **19**

Compound **19** (cryst, CH₂Cl₂/EtOAc) was synthesized in 87% yield from **18** using a method similar to that described for the preparation of **13**. Mp 262–265 °C. ¹H-NMR (CDCl₃) δ : 0.65 (3H, s), 1.12–2.08 (10H, m), 1.14 (3H, s), 2.17–2.50 (3H, m), 2.61 (1H, d, J = 12 Hz), 2.95 (3H, s), 3.30 (1H, dd, J = 4, 12 Hz), 5.82 (1H, d, J = 10 Hz), 5.85 (1H, d, J = 7 Hz), 6.25 (1H, d, J = 7 Hz), 6.98 (1H, d, J = 10 Hz), 7.20–7.65 (10H, m). IR (KBr): 3312, 2946, 1706, 1664, 1602 cm⁻¹. HR-MS m/z: calc for C₃₃H₃₈N₂O₃ (M+): 510.2883; found: 510.2891. Anal calc for C₃₃H₃₈N₂O₃·6/5H₂O: C, 74.46; H, 7.65; N, 5.26. Found: C, 74.43; H, 7.41; N, 4.98.

Biological activity

Preparation of 5α-reductase from human and rat prostates Human and rat prostates were minced into small pieces. The minced tissue was homogenized in approximately three tissue volumes of buffer A (20 mM potassium phosphate, pH 6.5, containing 0.32 M sucrose, 1 mM dithiothreitol, 50 μM

NADPH and 0.001% PMSF), first with a Polytron (Kinematica GmbH) and then with a Teflon/glass homogenizer. The homogenate was centrifuged at 140 000 g for 60 min and then the pellets were washed with approximately three tissue volumes of buffer A. The washed pellets were used as the 5α -reductase.

Assay of 5α -reductase inhibitory activity

The reaction solutions contained 1 µM [14C]testosterone, 1 mM dithiothreitol, 40 mM buffer (potassium phosphate, pH 6.5, for the rat enzyme; Tris-citrate, pH 5.5, for the human enzyme), prostatic particulate (0.2–1 mg protein) and 0.5 mM NADPH in a final volume of 0.5 mL. The samples were added to 5 μ L DMSO and the control tubes received the same volume of DMSO. The reactions were carried out for 10-30 min and were stopped with 2 mL ethyl acetate containing testosterone, 5adihydrotestosterone and androstenedione (10 µg each). After centrifugation at 1000 g for 5 min, the ethyl acetate phase (upper) was transferred to a tube and then evaporated under nitrogen to dryness. The steroids were taken up in 30 µL ethyl acetate and the solutions were applied to Whatman LK5DF or LK6DF silica plates, and the plates were developed in ethyl acetate/cyclohexane (1:1) at room temperature. The plates were air-dried, and the chromatography was repeated. The radioactivity profiles were determined by bioimage analyzer (Fuji Film Co, Ltd).

The testosterone 5α -reductase inhibitory activities against human enzyme seem to reflect approximately the inhibition of the type 2 enzyme [7]. On the other hand, isozyme type is uncertain for rat enzyme [8].

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